

# Attention-Like Deficit and Hyperactivity in a *Drosophila* Memory Mutant

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The primary function of a brain is to produce adaptive behavioral choices by selecting the right action at the right time. In humans, attention determines action selection as well as memory formation, whereas memories also guide which external stimuli should be attended to (Chun and Turk-Browne, 2007). The complex codependence of attention, memory, and action selection makes approaching the neurobiological basis of these interactions difficult in higher animals. Therefore, a successful reductionist approach is to turn to simpler systems for unraveling such complex biological problems. In a constantly changing environment, even simple animals have evolved attention-like processes to effectively filter incoming sensory stimuli. These processes can be studied in the fruit fly, *Drosophila melanogaster*, by a variety of behavioral and electrophysiological techniques. Recent work has shown that mutations affecting olfactory memory formation in *Drosophila* also produce distinct defects in visual attention-like behavior (van Swinderen, 2007; van Swinderen et al., 2009). In this study, we extend those results to describe visual attention-like defects in the *Drosophila* memory consolidation mutant *radish*<sup>1</sup>. In both behavioral and brain-recording assays, *radish* mutant flies consistently displayed responses characteristic of a reduced attention span, with more frequent perceptual alternations and more random behavior compared with wild-type flies. Some attention-like defects were successfully rescued by administering a drug commonly used to treat attention-deficit hyperactivity disorder in humans, methylphenidate. Our results suggest that a balance between persistence and flexibility is crucial for adaptive action selection in flies and that this balance requires *radish* gene function.

## Introduction

The fruit fly *Drosophila melanogaster* has been used for >30 years as a model system to study behavior in general (Vosshall, 2007) and learning and memory specifically (Keene and Waddell, 2007). More recently, investigations relating memory defects to attention-like processes have been initiated (van Swinderen, 2005, 2007; van Swinderen et al., 2009). Numerous *Drosophila* mutants defective in various aspects of memory formation represent a promising starting point to study the interactions between attention-like processes, memory, and action selection. For instance, a mutant might fail to show learning because it might not attend appropriately to relevant stimuli during memory formation, consolidation, or retrieval. Attention-like processes can be measured by the suppression of a competing stimulus, and such suppression effects should display quantifiable alternation dynamics that are reflected in the ongoing behavior of an animal. To provide insight into attention in animals, one approach is to measure short-term behavioral processes during perceptual tasks. However, short-term processes are rarely characterized in *Drosophila* learning mutants, for example, in which often a single

population performance index may conceal informative defects in attention-like behavior. As a case in point, recent work has shown that the learning mutants *dunce*<sup>1</sup> and *rutabaga*<sup>2080</sup> appear to be less distracted by competing visual stimuli compared with wild-type flies, in both behavioral and electrophysiological paradigms (van Swinderen, 2007). Using a recently developed optomotor maze (van Swinderen and Flores, 2007), in which walking flies make successive turn choices in response to visual underlying moving gratings, *dunce* and *rutabaga* mutants were also found to display significantly greater optomotor responses in the maze than did wild-type flies, leading to the hypothesis that higher optomotor scores predict defective suppression mechanisms characteristic of attention (van Swinderen, 2007). Interestingly, the *Drosophila radish*<sup>1</sup> mutation did not show a significant optomotor response in the maze under the same conditions that produced strong responses in *dunce*<sup>1</sup> and *rutabaga*<sup>2080</sup>.

A gene involved in anesthesia-resistant memory (ARM) (Folkers et al., 1993; Tully et al., 1994; Folkers et al., 2006), *radish* has been suggested to define a separate pathway to memory formation distinct from *dunce* and *rutabaga* (Isabel et al., 2004). Considering the opposing optomotor phenotypes of *radish*<sup>1</sup> and *dunce*<sup>1</sup>, a major question was raised: why do mutants defective in supposedly distinct odor memory consolidation pathways also show distinct visual behavior phenotypes in the optomotor maze? Because *radish* mutant flies are not blind (Gong et al., 1998), we hypothesized that the poor response of *radish*<sup>1</sup> to visual stimuli in the maze results from defective short-term processes relevant to attention rather than from defects in vision per se. To

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address this question, we approached attention in *radish*<sup>1</sup> and wild type by three different paradigms amenable to addressing short-term processes: individual choice behavior in the optomotor maze, torque behavior in the tethered flight arena, and brain activity in response to competing visual stimuli. In the process of characterizing *radish* mutant defects in our visual paradigms, we developed a simple approach for measuring “attention span” in fly brain activity.

We found that *radish* mutants are defective for a number of short-term processes relevant to selective attention. These include the following: altered optomotor responsiveness, reduced turn stereotypy, and increased distraction in a visual choice maze; periodic hyperactivity and reduced fixation time in the tethered flight arena; and less sustained response dynamics as measured by local field potential (LFP) recordings in the brain.

## Materials and Methods

**D. melanogaster strains and stocks.** Flies were cultured at 22°C, with 50–60% humidity, on a 12 h light/dark cycle on standard media. Wild-type flies are from the Canton S strain; the *radish*<sup>1</sup> mutant, learning and memory mutants, and select Gal4 and upstream activating sequence (UAS) strains were obtained from the Bloomington *Drosophila* Stock Center. Targeted expression of the tetanus toxin light chain (*tnt*) or the *ether-a-gogo* (*eag*) mutation was produced by crossing a strain homozygous for insertions of UAS-*tnt* on the X chromosome ( $w^z, P\{w^{+mC} = UAS-TeTxLC\}$ ) or the strain  $w^z, P\{w^{+mC} = UAS-eag^{\Delta 932}\}/Cyo$  to the strain *Th-Gal4-3* ( $w^{1118}, P\{w^{+mC} = UAS-TH-3\}$ , a tyrosine hydroxylase construct on the third chromosome). The *hs-rsh(161)* strain (Folkers et al., 2006) was obtained from Scott Waddell (University of Massachusetts, Worcester, MA). Only 2- to 7-d-old female flies were phenotyped, 1 d after having been anesthetized with cold air (flight arena and electrophysiology) or CO<sub>2</sub>.

**Optomotor maze and population responses.** The maze, described previously (van Swinderen and Flores, 2007), consists of 3-mm-wide paths grooved into a 28 × 19 × 1 cm acrylic slab placed groove-face down on a glass plate to form a closed horizontal choice platform. The platform was positioned 3 cm above an up-ended 19-inch flat-screen computer monitor (Mitsubishi Diamondpro 930), on which was displayed a green/black grating of 1 cm spatial resolution, moving at 3 Hz (or other frequencies, as specified). The maze was surrounded by a white box (a 30 × 50 × 30 cm container), unless specified otherwise, with an aperture directly above for filming. Flies collected by CO<sub>2</sub> anesthesia the day before an experiment were loaded ( $n = 25$ –30) into modified, disposable polyethylene “jumbo” transfer pipettes (Fisher Scientific), in which they were allowed to acclimatize 3 min in the dark before the pipette was inserted into the starting position of the maze. After running the choice maze (2–5 min) in a darkened room, the flies’ distribution among the nine collection tubes was scored as a weighted average ranging from –4 to +4. The optomotor index (OI) is the deviation of the weighted average from 0, the middle tube, in which positive scores indicate optomotor responses in the same direction of image motion. All statistics were *t* tests of experimental means, unless otherwise specified. Visual stimuli presented to flies running the maze were exactly as described previously (van Swinderen and Flores, 2007): 1 cm green/black gratings moving at 3 Hz (or other frequencies, as specified). For distraction experiments, we followed two distinct paradigms. We modified a paradigm described previously (van Swinderen and Flores, 2007) involving static cardboard bars presented to either side of the maze. Because this paradigm did not allow for efficient calibration of the distracter, we developed a more flexible paradigm, featuring liquid crystal display (LCD) monitors, one on either side of the maze. The LCD monitors were ~10 cm from each side of the maze, facing each other. The LCDs displayed a 25 × 5 cm white vertical bar on a red background; the bar was placed only on the LCD that was on the side opposite optomotor flow on the cathode ray tube (CRT) (with the other LCD displaying only red). The percentage of background red in the white bar was gradually increased (from 0 to

100%) to measure the distractive effects of the bar on the optomotor response. Phototactic responses to the distracter alone were also tested, and fly distributions in the maze were quantified the same way as for optomotor experiments. In a second distraction paradigm, we tested flies’ optomotor responses to competing objects (fields of 1 cm blue squares or green circles) moving in opposing directions on the CRT beneath the maze.

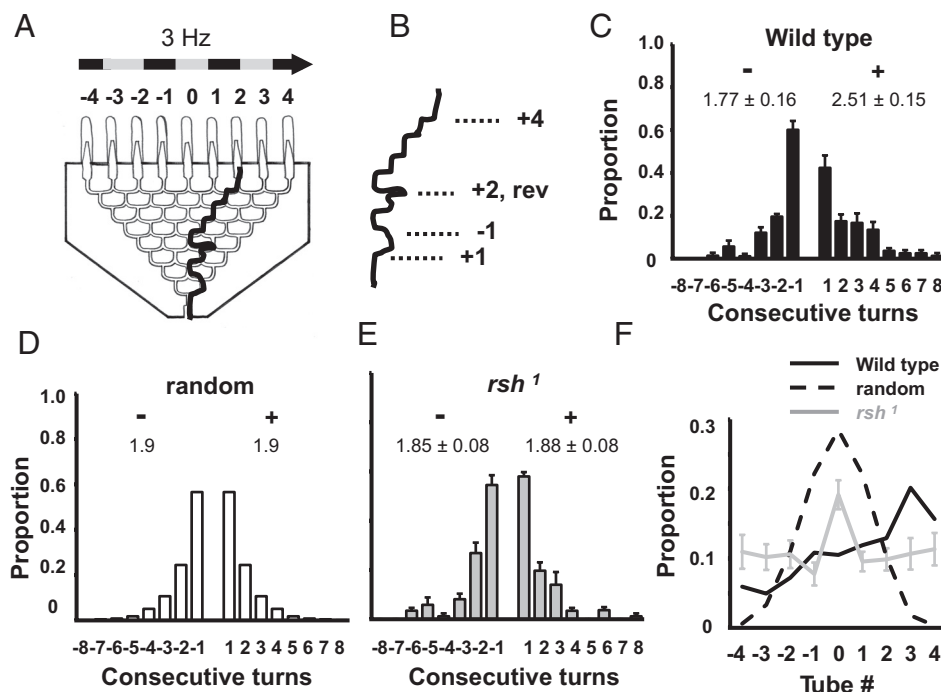
**Individual fly responses in the maze.** Playback from filmed experiments allowed individual fly choice behavior in the maze to be quantified. For a detailed analysis of optomotor behavior, the path followed by the first 10 flies to complete the maze was manually traced onto a template of the maze ( $n = 40$  per experiment, distributed among four separate mazes). At each of the eight successive choice points, the number of flies making “positive” or “negative” turns was tallied, providing a probability statistic for turns at each level. Occasionally, flies reversed direction, and these events were tallied as well according to the choice level where they occurred.

The number of consecutive turns in the same direction, before turning to the alternate direction or reversing, was tallied per fly as well, yielding a choice persistence index. Each fly could make a number of consecutive turns in either direction, up to a maximum of eight consecutive turns (which would necessarily position it in tubes –4 or +4). These data were summed for 10 flies per maze (40 in total) and averaged for a genotype. A weighted average of these eight possible categories was calculated for either direction (total 1 turn \* 1 + total 2 turns \* 2 + ... + total 8 turns \* 8), yielding a measure of choice persistence, which was normalized by the total number of turns. Perfect alternation behavior would thus yield a score of 1, random behavior would yield a score of 1.9 [based on a random model produced in MATLAB (MathWorks); data not shown], and significantly greater persistence scores indicate consistent turns in the same direction (stereotypy). In all of our individual fly analyses, we only quantified the behavior of the first 10 flies to complete each maze run.

**Turning behavior in the flight arena.** Flies were tethered as described previously (Maye et al., 2007) and tested the following day for flight behavior (for a video of the procedure, see Brembs, 2008). The duration of the experiments had to be confined to 6 min because *radish* mutants were reluctant to fly continuously in the arena. For the same reason, in some experiments [notably, for *hs-rsh(161)*], the number of animals is comparatively low. A featureless cylindrical arena was used to test baseline torque dynamics during uninterrupted 6 min flights, sampled at 20 Hz. In the closed-loop scenario, the setup was identical except that two pairs of opposing visual objects (upright and inverted Ts) were placed in the center of the four quadrants on the arena wall, and the speed of arena rotation was made inversely proportional to the yaw torque signal of the fly (Dill et al., 1993; Brembs and Heisenberg, 2000, 2001). Median fixation time for each strain was determined by durations of uninterrupted flight orientations centered on each of four 70° wedges (centered on each object).

**Activation of wild-type *radish*.** A wild-type *radish* transcript under the control of a heat-shock promoter, *rsh(161)* (Folkers et al., 2006), was activated in a *radish* mutant background by exposing flies (late-stage pupae or third-instar larvae) to 37° for 5 h.

**Electrophysiology.** Brain recordings were performed exactly as described previously (Nitz et al., 2002; van Swinderen, 2007). The recording site was 50–75 μm down from the center of the ocelli, with a reference electrode 100 μm into the top of the left eye. The recording is thus a voltage differential between these two sites, probably representing field effects produced by populations of mushroom body (MB) neurons near the central electrode (van Swinderen et al., 2009). Spectral analyses of brain activity data were performed in MATLAB by Fourier analysis of 24 overlapping sectors of the 360° rotating panorama. To determine significant selection/suppression of visual objects, we contrasted (by *t* test) the power of a bandpass-filtered signal for the six sectors (of 24 total) comprising one object (e.g., the cross in front of the fly) versus the six sectors comprising the opposing object (e.g., the square in front). A visual explanation of our responsiveness calculations is shown in supplemental



**Figure 1.** The optomotor maze paradigm. **A**, Maze. Arrow, Grating direction and velocity. Black trace, Filmed path of an individual fly. **B**, Each individual fly path is quantified for turning behavior. The number of successive turns in the same direction (+, with the moving grating; −, against the grating) is tallied per fly. Reversals of direction were also counted (rev). **C**, The normalized frequency of consecutive turn categories is plotted as a histogram for wild-type flies ( $n = 40$  flies; weighted average  $\pm$  SEM). The average value for either direction is indicated. **D**, Histogram for data created by a random model (50% turn probability at each choice level), with corresponding consecutive turn averages. **E**, Histogram and turn averages for *radish*<sup>1</sup> mutants ( $n = 40$  flies). **F**, Average  $\pm$  SEM distribution of flies among the nine collection tubes at the end of the maze ( $n = 8$  mazes of 25–30 flies for wild type and *radish*<sup>1</sup>) compared with a theoretical distribution for the random model.

Methods 1 (available at [www.jneurosci.org](http://www.jneurosci.org) as supplemental material). To determine alternation dynamics of 20–30 Hz activity in the ongoing attention paradigm, we calculated the log ratio of summed 20–30 Hz activity for successive 1.5 s epochs corresponding to either object sweeping in front of the fly. These ratios were then plotted to determine the duration of successively biased 20–30 Hz activity (in which activity was higher for one object for several cycles) until power alternated to the competing object. This duration is termed “alternation time” (AT). To quantify ongoing bias for either object, we summed AT values found within contiguous groups (“clumps”)  $>4$  and flanked by AT scores of 1. AT data from different flies were combined for all flies within a genotype (generally, all flies within a genotype revealed a similar number of such clumps for 400 s experiments) and plotted as frequency histograms. To determine the significance of such AT clumping, we shuffled (by permutations in MATLAB) the original log ratio data such that the temporal aspect of the data was lost, and then we recalculated AT values and resulting AT clump sizes. One thousand permutations of the data were performed to generate a frequency histogram for shuffled data. Significant differences between datasets was set at  $p < 0.05$  and determined by the Kolmogorov–Smirnov test for distributions. A visual explanation of our “attention span” calculations is shown in supplemental Methods 2 (available at [www.jneurosci.org](http://www.jneurosci.org) as supplemental material).

**Pharmacology.** Flies were treated with methylphenidate (MPH) (Sigma) mixing the drug (0.5 mg/ml) into regular fly food. Before feeding, flies were starved in empty vials for 2 h. Ingestion of the drug was confirmed by adding blue food coloring to the food and checking fly abdomens for blue color after feeding. Most flies had ingested drug-laced food by 3 h. Control flies were fed only blue food without drug. For electrophysiology experiments, tethered flies were provided with a morsel of the drug-laced food after  $\sim 2$  h starvation and observed to feed on it, with experiments starting 10 min to 1 h afterward. All flies prepared for electrophysiology were fed a morsel of food before testing in the arena, so these served as controls for the MPH-treated flies.

## Results

### Behavioral paradigms: optomotor maze

Wild-type flies respond to moving visual effects by turning in the direction of image motion (Heisenberg and Wolf, 1984) (an optomotor response, as measured by the OI; see Materials and Methods). In our optomotor maze paradigm, whereas wild-type Canton S flies display an average optomotor index of  $0.75 \pm 0.08$ , *radish* mutants showed diminished responsiveness to moving visuals in the maze (OI =  $0.10 \pm 0.12$  for green/black 1 cm grating moving at 3 Hz; see Materials and Methods) (van Swinderen, 2007). Because this *radish*<sup>1</sup> phenotype is not attributable to blindness [*radish* mutants can respond to visual objects in other conditions (Gong et al., 1998)], we wondered whether *radish* mutants might be characterized by distinct behavioral idiosyncrasies that interfered with choice behavior as they completed the maze. To measure short-term processes in the maze, we filmed and quantified individual behaviors as flies progressed through the eight-level choice maze (Fig. 1A). Turn stereotypy, defined as the average number of consecutive turns a fly made in the same direction (Fig. 1B) (see Materials and Methods), was tallied for 40 individuals. Wild-type flies display significant stereotypy in the direction of optomotor flow, making  $\sim 2.5$  consecutive turns in that direction compared with  $\sim 1.8$  consecutive turns against optomotor flow (Fig. 1C). This short-term hysteresis, or behavioral persistence, most likely contributes significantly to the positive optomotor score (van Swinderen and Flores, 2007). If turn behavior were random and independent across choice levels, one would see a symmetrical distribution resembling a binomial model, with  $\sim 1.9$  consecutive turns in the same direction (Fig. 1D). Interestingly, *radish* mutants behaved much like the binomial model, lacking any hysteresis across choice levels, with  $\sim 1.9$



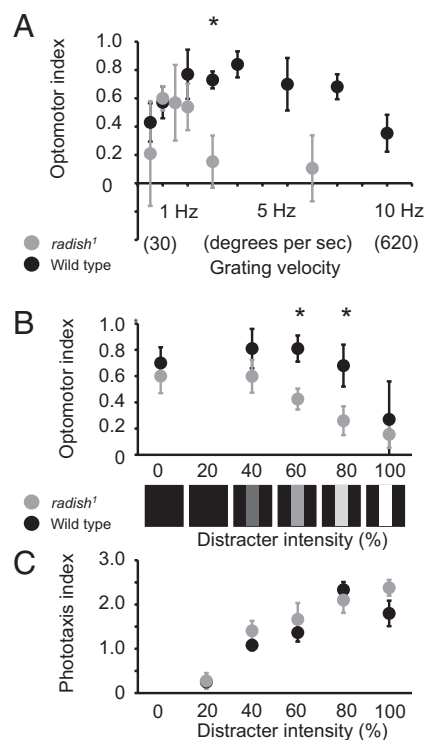
consecutive turns either with or against optomotor flow (Fig. 1E). The even distribution of *radish* animals across the nine end-points of the maze further suggested a defect in visual responsiveness in *radish* mutants (Fig. 1F).

Weak optomotor performance in the maze caused by the *radish* mutation was rescued by activating a wild-type *radish* transgene, *hs-rsh(161)*, on a mutant *radish* background. Interestingly, rescue required prolonged (5 h) activation of the *radish* gene during development. The heat-shock regimen (see Materials and Methods) used previously (Folkers et al., 2006) to rescue *radish*<sup>1</sup> for odor-learning defects in the adult was not sufficient for rescuing optomotor defects (before heat shock,  $OI = 0.09 \pm 0.15$ ; after heat shock,  $OI = 0.18 \pm 0.14$ ). However, *radish* transgene activation during the late pupal stage rescued optomotor responsiveness to our standard 3 Hz grating ( $OI = 0.55 \pm 0.12$ ), but activation during the third-instar larval stage did not ( $OI = 0.06 \pm 0.4$ ). The requirement of *radish* during development for normal optomotor responses resembles similar results found for *dunce* mutants (which display extremely high optomotor responsiveness) in which only *dunce* gene induction during development (but not adulthood) rescued *dunce* optomotor phenotypes (van Swinderen, 2007). Furthermore, *dunce* defects were also found to be rescued specifically at the late pupal stage (van Swinderen et al., 2009). These results suggest that wild-type optomotor responsiveness levels are dependent on cellular events that occur during a specific stage of brain development and that *dunce* [a cAMP phosphodiesterase (Davis, 2005)] and *radish* [a gene with no clear homology (Folkers et al., 2006)] may be required during the same pupal stage for the formation of circuits affecting optomotor responsiveness.

*Radish* mutants are not blind and respond to visual stimuli, as has been shown in other behavioral studies (Gong et al., 1998). Phototaxis appears to be normal in *radish* mutants when tested in response to a light source in our maze paradigm ( $OI = 2.4 \pm 0.15$  vs  $OI = 2.55 \pm 0.08$  for wild type). To fully explore whether *radish*<sup>1</sup> responds at all to moving gratings, we exposed the mutants to different grating velocities. Wild-type flies typically display a velocity curve with decreased responsiveness at very low (<1 Hz) or very high (>10 Hz) grating velocities (Fig. 2A). We were surprised to find that *radish* mutants displayed a shifted velocity curve compared with wild type, with peak responsiveness at low (~1 Hz) grating velocities but no response at 3 Hz (Fig. 2A), which is the grating velocity we used in our initial behavioral characterization (above). Although this shifted optomotor effect in *radish*<sup>1</sup> requires additional explanation, the fact that *radish*<sup>1</sup> does respond to slower moving gratings allowed us to test the mutant for distraction phenotypes.

### Behavioral paradigms: distraction in the maze

Distraction can be measured by the loss of optomotor responsiveness to a moving grating caused by competing visual objects (van Swinderen and Flores, 2007). To test *radish*<sup>1</sup> for distractibility, we used two different paradigms that presented competing objects in different ways. In the first paradigm, we modified a previously used method in which static bars presented to the side of the maze abolish optomotor responses to the moving grating displayed under the maze (van Swinderen and Flores, 2007). To better quantify distraction phenotypes, we replaced the static (cardboard) distracters of our previous paradigm with programmable displays on LCD screens and measured responsiveness to the grating in competition with increasingly salient distracters (see Materials and Methods). We found that *radish* mutants (re-

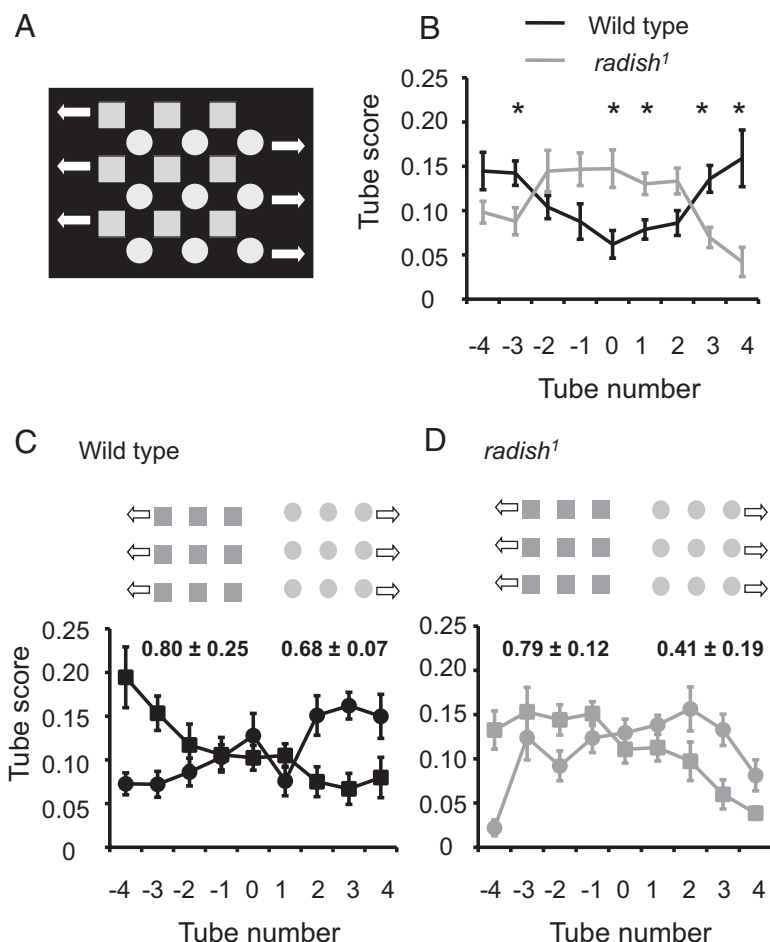


**Figure 2.** *radish*<sup>1</sup> optomotor responsiveness and distractibility. **A**, Optomotor responsiveness (means ± SEM;  $n = 8$  maze runs for each point) for wild type and *radish*<sup>1</sup> for different grating velocities. *radish* mutants do not respond to gratings moving 3 Hz or faster but do respond to more slowly moving gratings (1 and 2 Hz,  $p < 0.05$  by  $t$  test against zero). \* $p < 0.05$ , significantly different from wild type by  $t$  test of means. **B**, Optomotor responsiveness (means ± SEM;  $n = 8$  maze runs for each point) to a moving grating for wild type (at 3 Hz) and *radish*<sup>1</sup> (at 1 Hz; see above) in the presence of a visual distracter. A vertical white bar ( $5 \times 25$  cm) on a red background, positioned opposite of the direction of grating movement, was varied in intensity (percentage white; a schema is shown below the graph) to quantitatively test the distractibility of flies responding to the moving grating. \* $p < 0.05$ , significantly different from wild type by  $t$  test. **C**, Phototaxis responses of *radish*<sup>1</sup> and wild type in the maze to the distracter alone (the same schema shown above the graph).

sponding to a 1 Hz grating) lost optomotor responsiveness at lower levels of distracter intensity than wild type (Fig. 2B), although responsiveness to the distracter alone was the same for *radish*<sup>1</sup> and wild type (Fig. 2C).

To test the effect of distracting visuals in a different way, we presented them as competing moving objects on the CRT beneath the maze. Instead of the moving green/black grating, we displayed an intercalated field of green circles and blue squares moving slowly (~1 Hz) in opposite directions (Fig. 3A). Wild-type fly populations presented with this conflicting scene display a “U”-shaped distribution after completing the maze, suggesting that flies are following one or the other wide-field pattern for periods of time as they complete the maze (Fig. 3B, black line). In contrast, *radish* mutants display an “inverted U” distribution when presented with the competing optomotor stimuli (Fig. 3B, gray line). This suggests that, unlike wild type, *radish*<sup>1</sup> individuals do not follow one or the other wide-field stimulus for enough time to bias final fly distributions in the maze. *radish*<sup>1</sup> flies are nevertheless responsive to each wide-field stimulus presented independently, as are wild type (Fig. 3C,D).

Together, results from our visual competition paradigms as well as from filmed experiments over moving gratings suggest that the behavioral defects of *radish* mutants pertain to short-



**Figure 3.** Optomotor responsiveness to competing objects. **A**, A field of blue squares (1 cm wide) moving right to left at 1 Hz is intercalated with a field of green circles (1 cm diameter) moving left to right at 1 Hz (see Materials and Methods). **B**, Average  $\pm$  SEM distribution in the maze ( $n = 8$  maze runs) for wild type and *radish*<sup>1</sup> in response to the visual in **A**. \* $p < 0.05$ , significantly different proportion of flies by  $t$  test. **C**, Wild-type responses to the moving squares or circles presented individually, with average  $\pm$  SEM optomotor indices ( $n = 8$  maze runs). **D**, *radish*<sup>1</sup> responses to the moving squares or circles presented individually, with average  $\pm$  SEM optomotor indices ( $n = 8$  maze runs).

term choice processes (e.g., increased distraction, random alternation dynamics, and attention-like deficits). To pursue this hypothesis, we further characterized short-term behavioral processes in *radish* mutants using an entirely different visual paradigm, the flight arena.

#### Behavioral paradigms: flight arena

Suspended at a torque meter, flies spontaneously initiate turning maneuvers by modulating their wing beat amplitudes, resulting in measurable torque effects around the fly's vertical body axis (yaw torque). We used the yaw torque signal to allow the flies to control the angular position of visual patterns on the arena wall (Heisenberg and Wolf, 1984). In such a feedback setup, flies are thus able to report ongoing decision-making by their choice of flight direction.

Further supporting the notion that *radish* mutant flies are not visually impaired, the mutants show an optomotor response in the flight arena and are able to stabilize the rotation of the visual patterns (i.e., fly straight). In our setup, we allowed the flies to choose between flying toward a pair of upright Ts and a pair of inverted Ts. Previous research has shown that *radish* mutants can distinguish and learn to associate these two patterns with aversive stimuli (Gong et al., 1998), but when we analyzed the flight re-

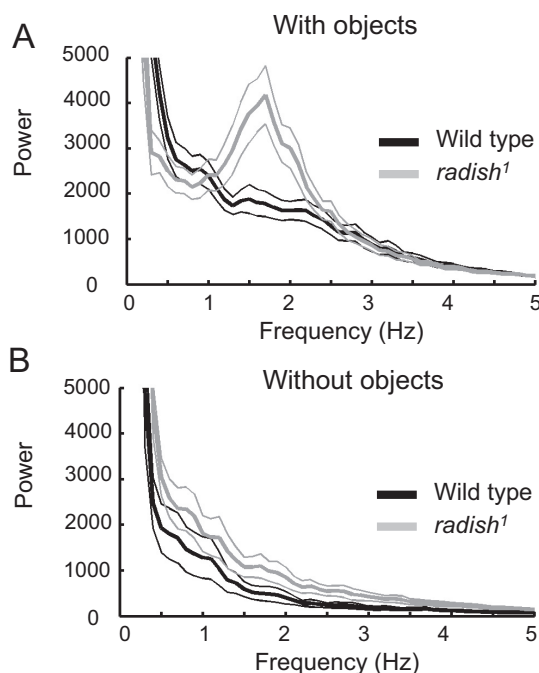
cordings of naive animals, we discovered that wild-type flies spend longer time periods directly in front of the patterns compared with *radish* mutants. The median time spent by *radish* mutants fixating on either pattern was significantly less than wild type: only 1.3 s compared with 3.0 s for wild-type flies ( $p < 0.01$  by Kruskal–Wallis nonparametric test for medians) (supplemental Fig. 1, available at [www.jneurosci.org](http://www.jneurosci.org) as supplemental material). We next analyzed the torque trace for any abnormalities that could explain this difference. Surprisingly, we found a strong  $\sim 1.6$  Hz peak (Fig. 4A, gray line) in the power spectrum of the *radish*<sup>1</sup> torque trace, whereas wild-type flies controlling the same visual stimuli did not show such oscillatory hyperactivity in their torque behavior (Fig. 4A, black line) (for sample torque traces, see supplemental Fig. 2, available at [www.jneurosci.org](http://www.jneurosci.org) as supplemental material). A different variant of *radish*<sup>1</sup>, *hs-rsh(161)* (Folkers et al., 2006), also showed this peak (supplemental Fig. 3, available at [www.jneurosci.org](http://www.jneurosci.org) as supplemental material), firmly establishing the mutated *radish* gene, rather than the genetic background, as the cause of this oscillatory hyperactivity.

To study whether this “fidgety” phenotype was generated endogenously or induced by the visual patterns, we analyzed the torque traces of flies recorded in the absence of any patterns. Without feedback or visual stimuli (i.e., open loop), we found that turning behavior in *radish* mutants was still characterized by some hyperactivity between 1 and 2 Hz. However, in this situation, hyperactivity was not

characterized by a single frequency peak, being instead more broadly distributed between 0.5 and 3 Hz (Fig. 4B, gray line). Wild-type torque behavior was again not strongly periodic without visual stimuli present (Fig. 4B, black line). These tethered flight data suggest that the presentation of competing visual stimuli induces oscillatory hyperactivity in *radish* mutants.

#### Brain recording paradigms: oscillations

We were curious as to whether the flight hyperactivity of *radish*<sup>1</sup> flies was indicative of more widespread activity effects in the brain. To determine this, we recorded LFPs using a tethered preparation that has been used previously to study brain activity in flies (Nitz et al., 2002). LFPs reflect the synchronous activity of populations of neurons, and these have been associated with visual (van Swinderen and Greenspan, 2003) as well as olfactory (Tanaka et al., 2009) stimuli in *Drosophila*. As in the flight arena, flies are tethered by their head and thorax but are free to move their legs and wings in this electrophysiology preparation (Fig. 5A). Two glass electrodes implanted into the brain and recording a voltage differential provide an ongoing measure of brain activity in the fly. Overall LFP activity in *radish* mutants was not significantly different from wild type, as evidenced by spectral analysis between 5 and 100



**Figure 4.** Tethered flight. **A**, Average  $\pm$  SEM power spectra between 0 and 5 Hz for wild-type (black line;  $n = 25$ ) and *radish*<sup>1</sup> (gray line;  $n = 24$ ) torque behavior in 6 min closed-loop flights with two distinct visual objects (see Materials and Methods). **B**, Average power spectra between 0 and 5 Hz for wild-type (black line;  $n = 25$ ) and *radish*<sup>1</sup> (gray line;  $n = 21$ ) torque behavior in 6 min open-loop flights without any visual landmarks.

Hz (data not shown). However, when we explored low LFP frequency domains (0–5 Hz) in *radish* mutants exposed to visual stimuli (van Swinderen and Greenspan, 2003), we did notice increased LFP activity between 1 and 2 Hz in most *radish* flies compared with wild type (Fig. 5B), although this was not significant on average for our sample.

In our brain recording preparation, our animals are not flying, and therefore they are not engaged in the behavioral alternations that produce torques in the flight paradigm. To determine whether the 1–2 Hz effect in brain activity might still be attributable to behavioral twitches, we inserted an electrode in the thorax to record movement (van Swinderen et al., 2004). Comparison of raw traces of brain and thoracic activity in *radish* mutants revealed that the 1–2 Hz brain oscillations were often sinusoidal, whereas the thoracic potentials were mostly bursts of spike-like activity, not necessarily synchronized to the brain events (supplemental Fig. 4, available at [www.jneurosci.org](http://www.jneurosci.org) as supplemental material). The source of the 1–2 Hz oscillation remains unclear; it may be widespread across the mutant's nervous system, as evidenced by a minor peak at similar frequency range from the thorax recording (supplemental Fig. 4, available at [www.jneurosci.org](http://www.jneurosci.org) as supplemental material). A recent report has shown a similar  $\sim 2$  Hz flight effect in wild-type flies lacking flight experience (Hesselberg and Lehmann, 2009). Together with our data, this suggests that *radish* mutants might be behaving like inexperienced flies in tethered flight.

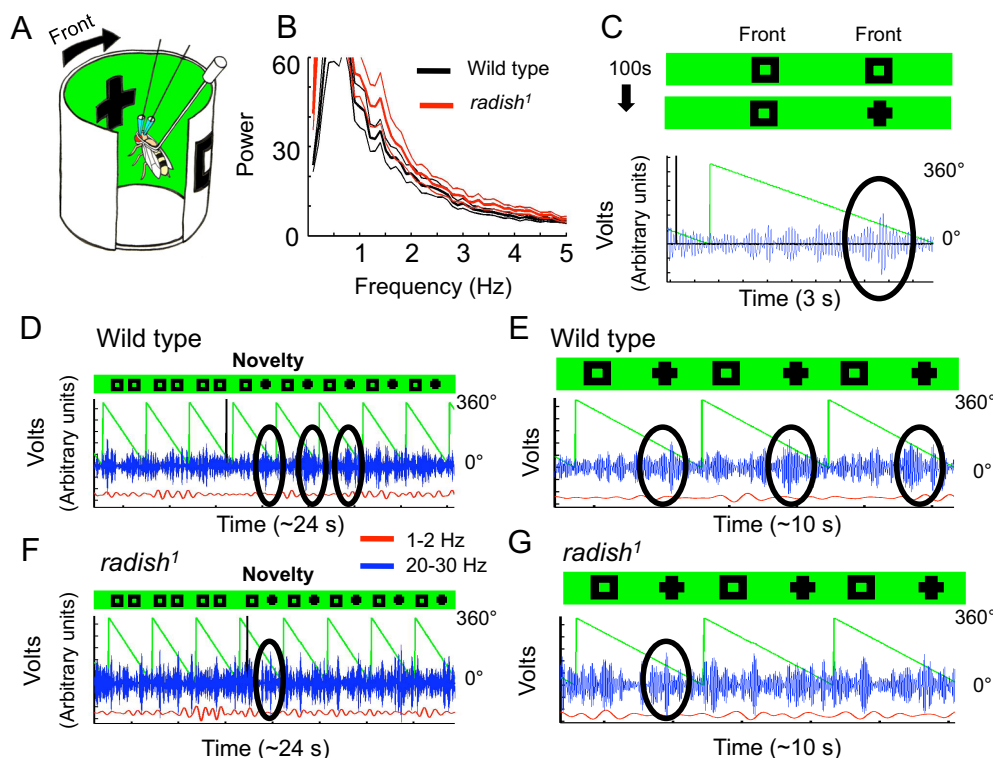
#### Brain recording paradigms: novelty detection

A more general explanation for the behavioral defects described so far for *radish*<sup>1</sup> would suggest that these mutants have an “attention deficit.” To address attention-like processes in the brain of *radish* mutants, we recorded LFPs while presenting competing

visual stimuli of variable salience. As described above, introducing competing visual objects to *radish* mutants in tethered flight resulted in a dramatically increased 1–2 Hz torque behavior, indicating that *radish* mutants are responding to the competing visuals by engaging in this strongly periodic behavior. Because the net consequence of this behavior is decreased time fixating either object (discussed above), this suggested an increased rate of perceptual alternations in *radish* mutants. We therefore proceeded to investigate brain responses to competing visual stimuli. Brain LFP activity in the 20–30 Hz range was shown previously to be associated with visual salience effects such as novelty, with wild-type flies showing a significant selection of visual novelty and simultaneous suppression of a competing non-novel object (van Swinderen and Greenspan, 2003; van Swinderen, 2007; van Swinderen et al., 2009). The learning and memory mutants *dunce*<sup>1</sup> and *rutabaga*<sup>2080</sup> are defective in their brain responses to visual novelty, and this is associated with increased optomotor behavior in the maze (van Swinderen, 2007). We probed whether brain LFP responses were also compromised in *radish* mutants because *radish*<sup>1</sup> performed poorly in the optomotor maze, were more distractible, and attended more briefly to visual stimuli in tethered flight relative to wild type.

In our visual novelty paradigm, flies are exposed to two identical squares rotating 180° apart around the fly for 100 s, and then one of the squares is changed to a cross (Fig. 5C). To investigate brain responsiveness to visual novelty, we bandpass filtered the LFP signal at 20–30 Hz and correlated amplitude of that frequency band to either competing object on the rotating panorama (Fig. 5C) (for a visual explanation, see supplemental Methods 1, available at [www.jneurosci.org](http://www.jneurosci.org) as supplemental material). Wild-type flies display a sustained increase in 20–30 Hz activity associated with a novel visual object, often lasting three to four cycles of the rotating panorama (Fig. 5D,E). In contrast, *radish* mutants did not appear to display a sustained response to visual novelty in the brain-recording paradigm (Fig. 5F,G). Indeed, averaging data for multiple flies ( $n = 14$ ) for the 10 s after a novelty transition suggested that *radish* mutants do not respond to visual novelty, for any LFP frequency domain between 10 and 40 Hz (Fig. 6A, left), whereas wild-type flies show a strong responsiveness to visual novelty (Fig. 6B, left) (van Swinderen, 2007).

Closer observation of the LFP signal suggested differences in the temporal dynamics of 20–30 Hz responses in *radish*<sup>1</sup> compared with wild type; the mutant appears to respond more often at various times of each panorama rotation without the suppression effects characteristic of wild-type flies (Fig. 5E,G). We wondered whether *radish* mutants might fail to detect novelty because of such defective attention-like dynamics, as also suggested by our behavioral experiments. To address this, we partitioned our data into successive 3 s epochs (approximately one panorama rotation, e.g., the individual green wedges in Fig. 5D,E), instead of averaging the response for the entire 10 s after a novelty transition as before. When analyzed on this finer temporal scale, wild-type flies show a sustained response to novelty for three to four successive rotations ( $\sim 9$ –12 s) of the competing images (Fig. 6B, right; and Fig. 5D,E in a sample fly). In contrast, when we looked at *radish*<sup>1</sup> responses on this finer time resolution, we found that the mutant flies did indeed respond to novelty but that this image-locked response only lasted on average 3 s, or only one exposure of the competing objects (Fig. 6A, right). This confirms that *radish* mutants can see and discriminate the objects and suggests that the defect here lies not in the perception of visual stimuli but rather in the duration of perception before an inter-



**Figure 5.** *radish*<sup>1</sup> brain recordings. **A**, Arena setup. Visual objects rotate around the fly clockwise with a period of 3 s. **B**, Average  $\pm$  SEM power spectrum of wild-type and *radish*<sup>1</sup> brain activity between 0 and 5 Hz ( $n = 14$  flies for both genotypes). The large peaks below 1 Hz (off scale) represents responses to the visual objects rotating around the fly at 0.33 Hz. **C**, Flies were exposed for 100 s to two identical squares before one of the squares changed to a cross. The amplitude (or power) of brain LFPs bandpass filtered at 20–30 Hz was correlated with the position of either competing object as it swept in front of the fly following a novelty transition. Data for a sample wild-type fly are shown. The black vertical line indicates when the square changed to a cross. **D**, Sample 20–30 and 1–2 Hz bandpass-filtered wild-type data for several rotations of the visual panorama before and after a novelty transition. Circled epochs indicate successive 20–30 Hz responses to the novel cross. **E**, Enlarged section of the novelty response in **D**. **F**, Sample 20–30 and 1–2 Hz bandpass-filtered *radish*<sup>1</sup> data for several rotations of the visual panorama before and after a novelty transition. **G**, Enlarged section of the novelty response in **F**. The response to the novel cross (circled) does not recur selectively, as it does in wild type.

nally generated switch occurs. Further confirming that the defect in this memory mutant is not at the level of simple visual processing, LFP responses to the same visual objects presented singly were not defective in *radish* mutants (Fig. 6C). Rather, it seems to be the introduction of salient visual competition that distracts the mutant sooner than wild type.

### Brain recording paradigms: attention span

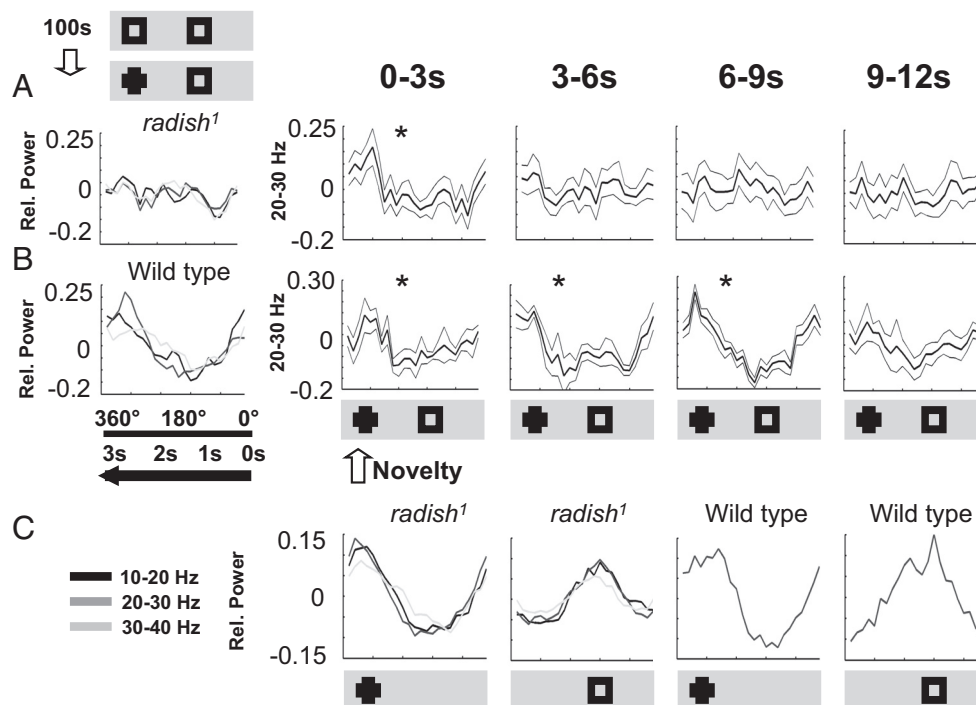
A central characteristic of attention is that it alternates among competing percepts at a characteristic tempo (Carter and Pettigrew, 2003; Brascamp et al., 2005). If the 20–30 Hz response to novelty reflects attention-like processes, then the abbreviated selective response to novelty in *radish*<sup>1</sup> may be indicative of a shortened attention span in this mutant. Furthermore, if 20–30 Hz responses to visual stimuli do indeed describe an attention-like process, then these should be continuously alternating among competing percepts even without experimentally imposed novelty. To study this possibility, and with a view to better understand *radish*<sup>1</sup> defects, we exposed flies to the same two distinct objects continuously, without novelty, while recording LFPs from their brains. We then applied some simple calculations to quantify alternation tempos in the brain LFP (supplemental Methods 2, available at [www.jneurosci.org](http://www.jneurosci.org) as supplemental material).

Wild-type flies and *radish* mutants were exposed to two objects in open loop (a square and a cross) simultaneously, as before (Fig. 5), but for extended epochs (400–800 s per fly) (Fig. 7). For

each recording, 20–30 Hz power was summed for the consecutive 1.5 s periods when either object was in the frontal hemifield of the fly (Fig. 7A, black and gray bars). Thus, for every full 3 s rotation of the panorama, two 20–30 Hz values were calculated, one for either object. The log ratios of these two 20–30 Hz values are plotted for a sample wild-type fly in Figure 7B, for 60 successive cycles of the panorama rotation. The 20–30 Hz activity often appeared to alternate in power between objects: when power for one object was high, it was then low for the other object during that cycle (Fig. 7B). At times (or in the special case of salience relating to novelty, as above) such partitioning of 20–30 Hz power was biased toward one object for several successive cycles before alternating. Timings (in rotation cycles) between such alternations, termed here as AT (alteration time), were tallied (Fig. 7B, gray and black numbers) and visualized by a histogram (Fig. 7C, for the same sample fly). Noticing that wild-type AT data looked “clumped” (i.e., epochs when 20–30 Hz was biased to one or the other object for successive cycles; there are 5 such clumps in Fig. 7C) (supplemental Methods, available at [www.jneurosci.org](http://www.jneurosci.org) as supplemental material), we quantified this LFP bias (labeled  $\Sigma$  AT) in the fly brain activity by summing the duration (in total rotation cycles) of each AT clump (Fig. 7C, corresponding numbers in the right column). Combined data from eight wild-type flies revealed a frequency histogram with a characteristic shape resembling a gamma distribution (Fig. 8A).

To determine the significance of this distribution of wild-type LFP alternation dynamics in response to two competing visual





**Figure 6.** Average responses to visual novelty in *radish*<sup>1</sup> and wild type. **A**, *radish*<sup>1</sup>. Left, Average LFP activity for the 10 s after a novelty transition for three frequency domains (10–20, black Hz; 20–30 Hz, dark gray; 30–40 Hz, light gray) for *radish* mutants ( $n = 14$  flies). Right, The same 20–30 Hz *radish*<sup>1</sup> data as summarized in the left but partitioned into successive 3 s epochs after a novelty transition ( $*p < 0.05$ , significant response) (supplemental Methods 1, available at [www.jneurosci.org](http://www.jneurosci.org) as supplemental material). **B**, Wild type. Left, Average LFP activity for the 10 s after a novelty transition was calculated for three frequency domains;  $n = 6$  flies. The direction of panorama flow is indicated. Right, The same 20–30 Hz wild-type data as summarized in the left but partitioned into successive 3 s epochs after a novelty transition [ $*p < 0.05$ , significant response (data are from van Swinderen, 2007)]. **C**, Average LFP responses to each of the two visual objects presented individually to *radish* mutants ( $n = 14$ ), for the three frequency domains indicated. Average wild-type 20–30 Hz responses to the same individual objects are shown for comparison (van Swinderen, 2007) ( $n = 6$ ).

objects, we contrasted our experimental data to shuffled data derived from the same experiments. In this way, any temporal relation between 20–30 Hz values measured for successive cycles of image presentation would be lost. Shuffling of the data indeed revealed a loss of clumping, or fewer contiguous epochs of biased 20–30 Hz activity (Fig. 7D). Tallying this bias ( $\Sigma$  AT) by a frequency histogram revealed a significantly different distribution compared with nonshuffled data ( $p < 0.05$ , Kolmogorov–Smirnov test for distributions), which was more characteristic of a binomial distribution (Fig. 8B).

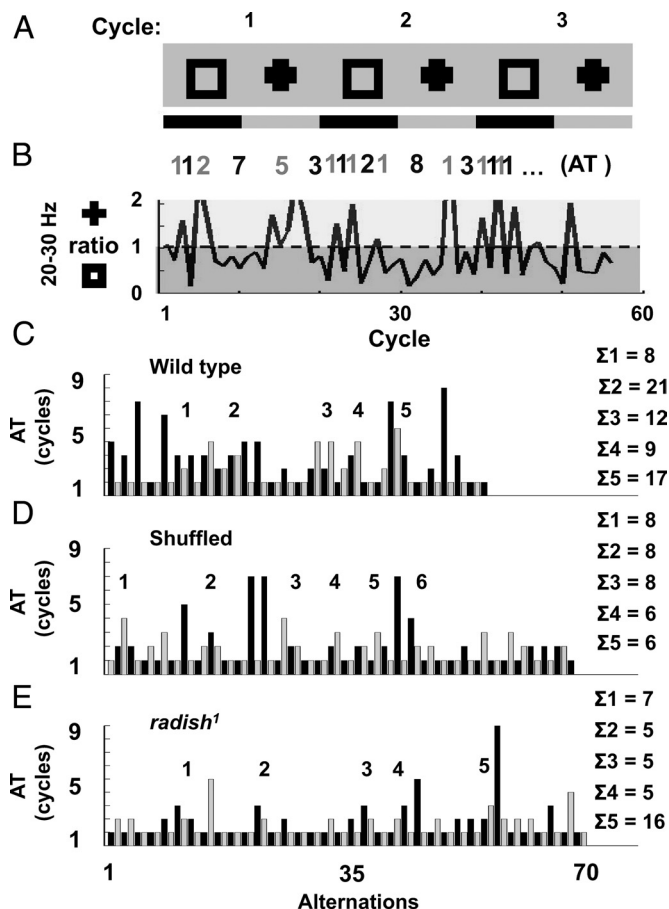
Finally, we applied the same methodology to *radish*<sup>1</sup> and found that these mutants displayed brain LFP alternation statistics that were not significantly different from shuffled wild-type data (Figs. 7E, 8C) ( $p = 0.85$  by the Kolmogorov–Smirnov test for distributions). This suggests that *radish*<sup>1</sup> brain activity is alternating randomly between the two visual percepts (the cross and the square), without the sustained “attention” across successive cycles characteristic of wild-type flies. Curiously, heat-shock of *hs-rsh(161)* failed to rescue this alternation defect in *radish* mutants but did result in improved novelty detection (supplemental Fig. 5, available at [www.jneurosci.org](http://www.jneurosci.org) as supplemental material).

### MPH effects

Given that we found a deficit in attention-like processes and a well defined behavioral hyperactivity in the *radish* mutant flies, we wondered whether drugs commonly used in humans to treat similar symptoms in perceptual timing and hyperactivity could also rescue analogous defects in flies. In humans, amphetamines are known to affect attention in a variety of ways, depending on

dosage and the specific drug used. In previous studies, we have shown that methamphetamines can increase optomotor responsiveness (van Swinderen and Flores, 2007) as well as modulate LFP responses to visual stimuli (Andreatic et al., 2005). Here, we asked specifically whether a related drug, MPH (“Ritalin”), could rescue *radish*<sup>1</sup> defects in the optomotor maze and recording arena. MPH treatment, targeting the dopaminergic system (Iversen and Iversen, 2007), is typically administered to patients with attention deficit and hyperactivity disorder (ADHD). To first test the effect of MPH on wild-type flies, we fed starved animals 0.5 mg/ml of drug mixed in fly food. Testing in the optomotor maze after feeding resulted in an increased optomotor response, but this became significant only after 3 h since the initial feeding (Fig. 9A, 3 h). Such an increase in responsiveness is consistent with previous results for acute methamphetamine treatment in flies (van Swinderen and Flores, 2007) and also consistent with the time course of MPH treatment in humans (Pelham et al., 1999). A full day of chronic MPH exposure further increased the wild-type response (Fig. 9A, 24 h), although flies appeared to be slower and lethargic (data not shown). For this population exposed to a chronic treatment of MPH, only 1–2 h off the drug and onto regular food completely reset optomotor responsiveness to normal levels (Fig. 9A, Recovery), suggesting that effects on the brain were transient. We decided to focus on acute MPH treatment (2–4 h) to test for phenotypic rescue in *radish* mutants exposed to black/green gratings moving at 3 Hz (where *radish*<sup>1</sup> showed no responsiveness). Remarkably, MPH treatment significantly increased *radish*<sup>1</sup> optomotor performance from zero to wild-type levels (Fig. 9B, *radish*). When we explored the generality of this effect (increased optomotor re-





**Figure 7.** Attention-like bias. **A**, Opposing visual stimuli (a square and a cross, 180° apart) rotate around the fly at 3 s per cycle. Each object is thus in front of the fly for 1.5 s, for which summed 20–30 Hz activity is separately calculated (during the epochs symbolized by the black and gray bars). **B**, Log ratio of summed 20–30 Hz activity plotted for successive cycles of image rotation in a sample wild-type fly. AT, Alteration time, or the duration (in cycles) when the ratio is biased in succession for one of the objects before alternating, indicated numerically above the graph. **C**, Successive AT values plotted as a time series histogram in the same sample wild-type fly. The size of five contiguous AT groupings (supplemental Methods 2, available at [www.jneurosci.org](http://www.jneurosci.org) as supplemental material) is tallied in a column on the right ( $\Sigma$  AT). **D**, Shuffled data from the same wild-type fly, replotted as a time series histogram with five tallied AT groups ( $\Sigma$  AT) shown on the right. **E**, The same analysis performed on data from the *radish*<sup>1</sup> mutant.

sponsiveness after drug treatment), we were surprised to find that acute MPH exposure did not alter optomotor responsiveness in a group of other learning and memory mutants (Fig. 9B), including *amnesiac*<sup>1</sup>, *rutabaga*<sup>2080</sup>, and *dunce*<sup>1</sup>.

MPH is believed to affect dopamine signaling by targeting recycling of the neurotransmitter at the synapse (Iversen and Iversen, 2007). To test the involvement of dopamine in MPH-mediated optomotor effects, we treated two classes of flies with genetically altered dopamine function (Friggeri-Grelin et al., 2003): *Th-Gal4/UAS-tnt* (in which dopaminergic neurons have been constitutively silenced) and *Th-Gal4/UAS-eag* [in which excitability of dopaminergic neurons was, in principle, increased (Broughton et al., 2004)]. Previous work has shown that either manipulation abolished optomotor responsiveness under our standard maze conditions, suggesting the requirement of a balanced dopamine environment for optomotor performance in the maze (van Swinderen and Flores, 2007). If MPH modulates dopamine levels in flies, then the presumed absence of dopamine in *Th-Gal4/UAS-tnt* mutants should block the effects of the drug.

Indeed, the drug failed to rescue optomotor responsiveness in *Th-Gal4/UAS-tnt* flies (Fig. 9B). In contrast, *Th-Gal4/UAS-eag* flies, in which dopamine function is altered but presumably not absent, showed increased optomotor responsiveness after drug treatment, like wild type and *radish*<sup>1</sup>.

We proceeded to investigate brain responses in *radish* mutants treated with acute MPH. A spectral analysis of LFP activity in the brain revealed that the 1–2 Hz oscillation characteristic of *radish* mutants was gone in animals treated with the drug compared with the same animals before treatment (Fig. 9C) (supplemental Fig. 6, available at [www.jneurosci.org](http://www.jneurosci.org) as supplemental material) ( $n = 5$ ;  $p < 0.05$  by paired  $t$  test comparing the average 1.4–1.7 Hz frequency domain before and after treatment). We then addressed effects on selective attention more specifically by means of our visual competition paradigm (as in Fig. 7). MPH did not significantly change the ongoing attention span defect of *radish* mutants, with alternations still appearing random in our sample (supplemental Fig. 5, available at [www.jneurosci.org](http://www.jneurosci.org) as supplemental material). However, when we introduced novelty salience into this paradigm (as in Figs. 5, 6), MPH exposure improved the 20–30 Hz response in *radish* mutants by producing an increase in power assigned to the novel stimulus (selection) and a decrease in power to the non-novel stimulus (suppression), resembling wild-type responsiveness to visual novelty in both amplitude and temporal dynamics (Fig. 9D) (supplemental Fig. 6, available at [www.jneurosci.org](http://www.jneurosci.org) as supplemental material). Acute exposure to MPH in wild-type flies did not further increase the LFP effect (Fig. 9D).

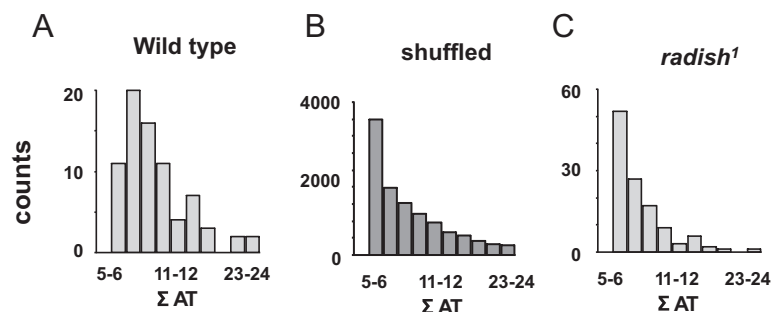
## Discussion

It is increasingly apparent that many classical *Drosophila* learning and memory mutants are also defective in short-term processes relevant to selective attention. Previous studies have shown that short-term memory as well as long-term memory mutants display attention-like defects (van Swinderen, 2007; van Swinderen et al., 2009), and the current study reveals *radish* mutants to be defective as well, albeit with distinctly different symptoms. The *Drosophila* mutants *dunce*<sup>1</sup>, *rutabaga*<sup>2080</sup>, and *radish*<sup>1</sup> share olfactory memory defects but differ conspicuously for short-term processes relevant to visual attention. Whereas the more persistent optomotor behavior of *dunce*<sup>1</sup> and *rutabaga*<sup>2080</sup>, both affecting the cAMP-associated pathways (Davis et al., 1995), are reminiscent of the persistent preoccupation of some patients afflicted with autism, the phenotype of *radish* mutant flies described here is similar to some of the symptoms of patients with ADHD.

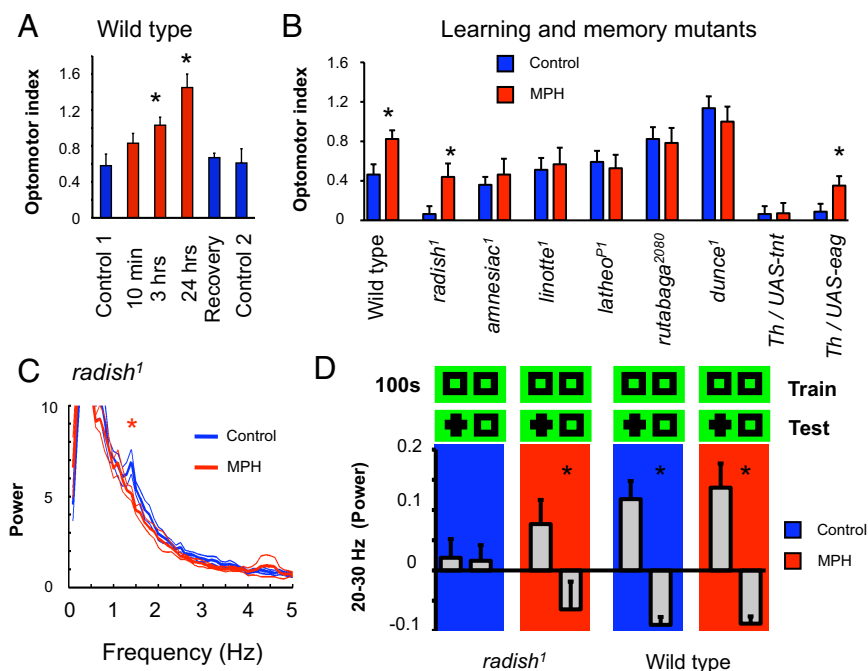
The *Drosophila radish* gene encodes a protein required for ARM (Folkers et al., 1993, 2006). Without a functional form of this protein, flies are unable to preserve memories through cold-induced anesthesia. Originally described as a step to long-term memory formation in a cAMP pathway (Tully et al., 1994), *radish* has more recently been proposed to be involved in a distinct parallel network modulating long-term memory formation (Isabel et al., 2004). The proposed separation of *radish* circuits from cAMP-associated pathways to memory formation is consistent with our MPH results, in which none of the cAMP pathway mutants showed altered performance in the maze after drug treatment, whereas *radish*<sup>1</sup> (and wild type) did. Because MPH is thought to target dopamine function in mammals (Iversen and Iversen, 2007), this suggests that dopaminergic modulation of attention-like phenotypes is blocked in mutants affecting cAMP

signaling but not blocked in *radish*<sup>1</sup>. Interestingly, although representing a separate molecular network, *radish* appears to be strongly expressed in the MBs (Folkers et al., 2006), the same memory-associated structures expressing *dunce*, *rutabaga*, and other cAMP-related molecules (Davis, 2005). Our finding that MPH treatment does not improve optomotor behavior for mutants in which dopaminergic neurons have been silenced suggests that dopamine input to the MBs (Schwaerzel et al., 2003) is upstream of cAMP-signaling neurons in the MBs but parallel to *radish*-expressing neurons in the MBs. Our results therefore predict that *radish* and *dunce* effects involve different neuronal populations.

*radish* has been implicated in appetitive learning (Krashes and Waddell, 2008), so its role in memory formation is not necessarily restricted to ARM and may instead relate to more general attention-like processes. One intriguing possibility supported by results from our brain recording and behavioral experiments is that *radish* circuits might be involved in modulating the tempo of stimulus selection and suppression in the fly brain. We show in this study that *radish* mutants fail to respond to visual novelty for the typical three to four stimulus exposures, which are the number of exposures for a characteristic novelty response in wild-type flies in our paradigm (van Swinderen, 2007; van Swinderen et al., 2009). The *radish* mutant discriminates visual novelty by a characteristic selection/suppression response but only responds for one exposure. Perhaps as a consequence of such abbreviated perceptual selection, *radish*<sup>1</sup> LFP alternations between competing visuals appear random over extended presentations. In contrast, wild-type flies displayed sustained biases in LFP responses, resulting in gamma-like distributions of response alternations. Such distribution shapes have also been observed in studies of perceptual rivalries in humans (Carter and Pettigrew, 2003). The significantly abbreviated temporal characteristics of *radish*<sup>1</sup> responsiveness were reflected in two different behavioral paradigms: the optomotor maze and tethered flight. Together, our behavioral and electrophysiological results suggest that alternation processes in *radish* mutants are defective: whereas wild-type flies showed some persistence or hysteresis between alternations in behavior or brain activity, *radish* mutants alternated too quickly or even in an oscillatory manner when presented with competing visual stimuli. In contrast, at least behaviorally, *dunce* and *rutabaga* mutants exhibit an opposite phenotype; both display persistent choice behavior in the optomotor maze and both are less distracted by competing visuals than wild type (van Swinderen, 2007; van Swinderen et al., 2009).



**Figure 8.** Distribution of attention-like bias. **A**, Frequency distribution of AT data for eight wild-type flies exposed to the two competing objects, as in Figure 7.  $\Sigma$  AT, A measure of attention-like bias, is the sum of AT values within each grouping (as indicated in Fig. 7C–E). **B**, Frequency distribution for eight sets of shuffled wild-type data. **C**, Frequency distribution for summed AT data from 14 *radish* flies.



**Figure 9.** MPH treatment. **A**, Optomotor responsiveness in wild-type flies treated with 0.5 mg/ml MPH (red histograms). Starved flies were transferred to drug-laced food and allowed to feed for 10 min, 3 h, or 24 h. Control flies were similarly transferred but to food without drug and tested 10 min later (Control1) or the next day (Control2). Flies chronically exposed to drug for 24 h were transferred back to normal food for 1–2 h and tested (Recovery). \* $p < 0.05$ , significantly different from controls by  $t$  test.  $n = 8$  runs of 25–30 flies for each experiment. **B**, MPH (0.5 mg/ml) was administered acutely (2–4 h feeding on drug-laced food) to a panel of learning and memory mutants, as well as to flies with altered dopamine function [*Th-Gal4* (Friggi-Grelin et al., 2003)  $\times$  *UAS-tnt*, wherein dopaminergic neurons are silenced, or  $\times$  *UAS-eag* <sup>$\Delta$ 32</sup>, in which dopaminergic neurons are activated]. \* $p < 0.05$ , significantly different from controls (red vs blue bars) by  $t$  test.  $n = 8$  runs of 25–30 flies for each experiment. **C**, Spectral analysis (0–5 Hz) of LFP activity in the brains of *radish* mutants treated to acute MPH (red line). Blue line, The same flies before treatment. Data are averages of z-scored spectrograms ( $n = 5$  flies). \* $p < 0.05$ , significantly different within 0.3 Hz band range surrounding the peak, by  $t$  test. **D**, The effect of MPH on 20–30 Hz responsiveness ( $\pm$  SEM) to visual novelty in *radish*<sup>1</sup> and wild-type flies. MPH feeding (red box) resulted in a significant novelty response in *radish* mutants ( $n = 5$  flies), whereas *radish* mutants fed without the drug (blue box) show no response to novelty ( $n = 14$ ). Wild-type responses to visual novelty were similar with and without MPH ( $n = 4$  and 6 flies, respectively). \* $p < 0.05$  by  $t$  test of means for either competing object. A visual explanation of how these data were calculated is in supplemental Methods 1 (available at [www.jneurosci.org](http://www.jneurosci.org) as supplemental material).

Attaining the right balance between persistence and flexibility is a crucial feature of adaptive behavior, because it reflects the balance between exploration and exploitation of natural resources. It is tempting to speculate that *radish* and *dunce/rutabaga* may constitute the two respective extremes of this balance. Recent work investigating torque behavior of wild-type flies

(similar to our shorter experiments here) has shown that, during extended flights, the occurrence of turning maneuvers can be described by a Lévi distribution (Maye et al., 2007). Such distributions of behavioral output, seen in foraging behavior in many animals, are characteristically long-tailed. This means that animals may occasionally persist with one behavioral choice for unusually long, but most often choices alternate at a more regular, normally distributed rate. The advantage of allowing for occasional long forays into one direction is presumably to chance on a new resource away from the proximal search space. Such behavior has been found to be ecologically advantageous, but mechanisms driving such alternation tendencies have not been documented in the *Drosophila* brain. One interpretation of our results is that the mushroom body circuits defined by *dunce/rutabaga/radish* expression are involved in establishing the balance between persistence and flexibility [i.e., the explore/exploit dilemma (Daw et al., 2006)]. A separate set of results has independently also arrived at a similar conclusion, suggesting that the mushroom bodies could be involved in maintaining a period of behavioral flexibility (i.e., attention-like processes) before a longer-term transition to habit formation or motor learning (Brembs, 2009).

The most intriguing open question following from our results is how the divergent attention-like effects in the classical learning mutants can be related to their converging memory defects. Of course, there may not be a connection at all. The common learning phenotype is restricted to classical olfactory learning; in visual learning, *radish*<sup>1</sup> appears to perform reasonably well, whereas *dunce* and *rutabaga* mutants fail (Gong et al., 1998; Brembs and Plendl, 2008). Moreover, the olfactory memory phenotypes of these mutants are rescued acutely (McGuire et al., 2003; Folkers et al., 2006), whereas the short-term defects described here and elsewhere (van Swinderen, 2007; van Swinderen et al., 2009) can only be rescued by expressing the wild-type gene during development and not in the adult. Surprisingly, our MPH results suggest that some of these short-term deficiencies (most likely caused by developmental defects in the brain) can be acutely compensated via alternate pathways modulating dopamine function.

Although distinct functions at this point cannot be excluded, it nevertheless remains tantalizing to find attention-like defects in every *Drosophila* memory mutant investigated so far: *dunce*<sup>1</sup> and *rutabaga*<sup>2080</sup> (van Swinderen, 2007), long-term memory mutants (van Swinderen et al., 2009), and *radish*<sup>1</sup> (this study). This may not be surprising: early studies already pointed to habituation and sensitization defects in some of these mutants (Duerr and Quinn, 1982), which may relate to the attention-like phenotypes of our study. We suggest that *radish*<sup>1</sup> may be defective in memory consolidation because of a previously overlooked defect in tempo of attention-like alternations. Reduced hysteresis in brain activity and behavior in the *radish* mutant, often amounting to random choice behavior, and especially a 1–2 Hz oscillation in brain activity and behavior, might interfere with the reconstitution of stereotypical patterns of neural activity representing a memory after anesthesia. Rather than calling for a reevaluation of the role of *radish* in memory formation, these results strengthen the view that ARM circuits act as a gating mechanism for memory formation (Isabel et al., 2004). By periodically changing perception from one stimulus to the next, attention mechanisms would thus play a role in gating memory formation. One challenge now is to understand how suppression mechanisms, modulated by *dunce/rutabaga* circuits (van Swinderen, 2007), interact with the temporal mechanisms modulated by *radish*

(this study) to control attention processes, action selection, and eventual memory formation.

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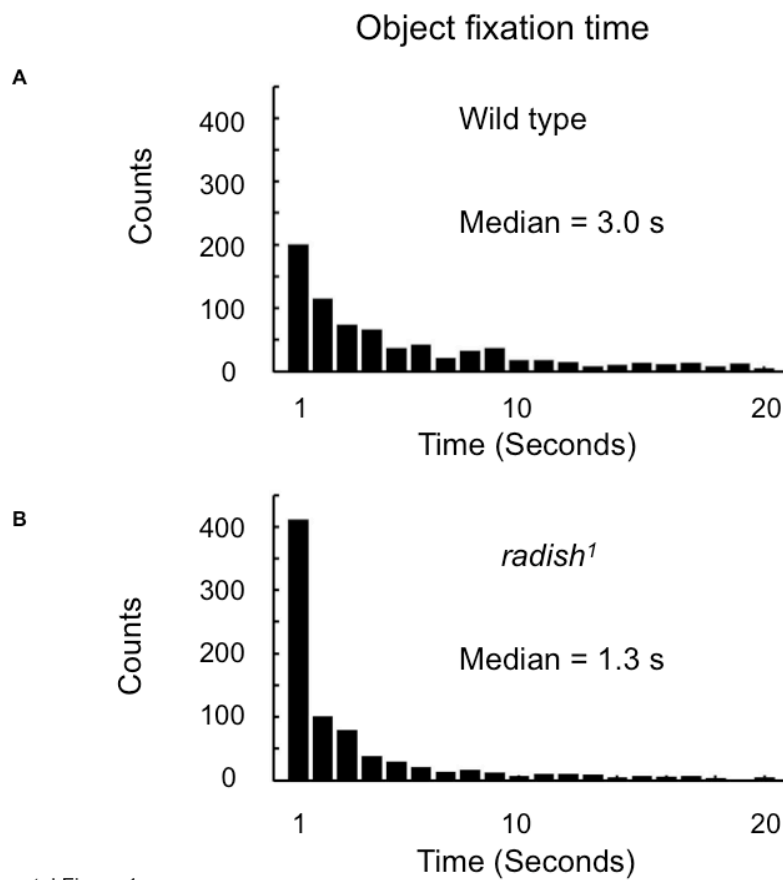
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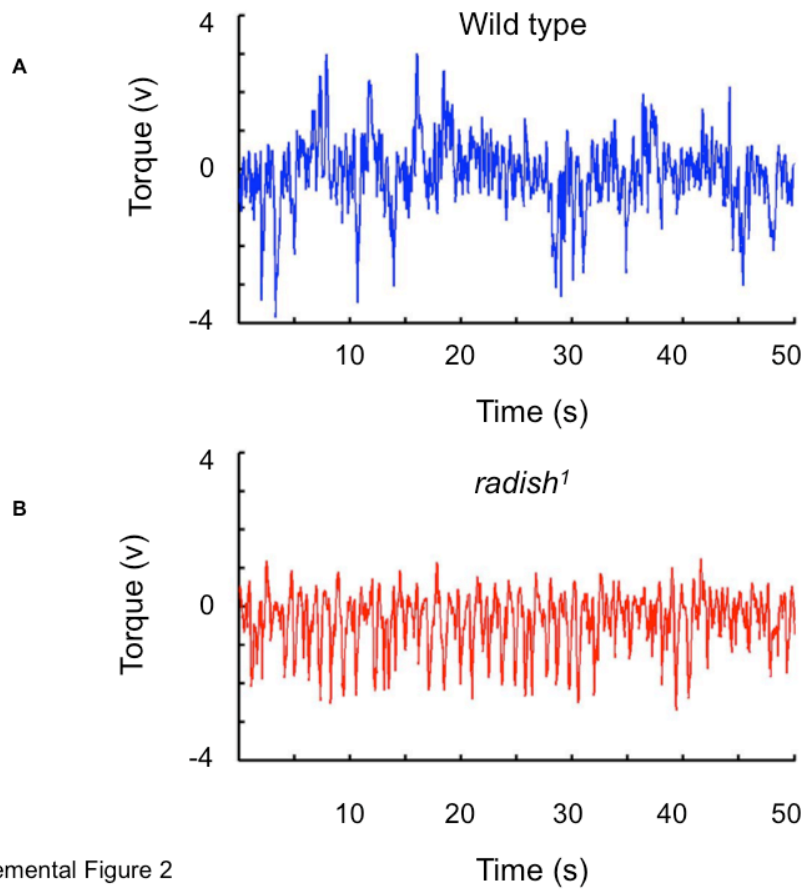
## Supplemental Material

**Supplemental Figure 1.** Frequency histogram for fixation times in the tethered flight arena (see Methods). Counts for each time bin are combined for all flies within a genotype; median fixation time is indicated. **A** wild type  $n=25$  **B** *radish*<sup>1</sup>  $n=24$ .



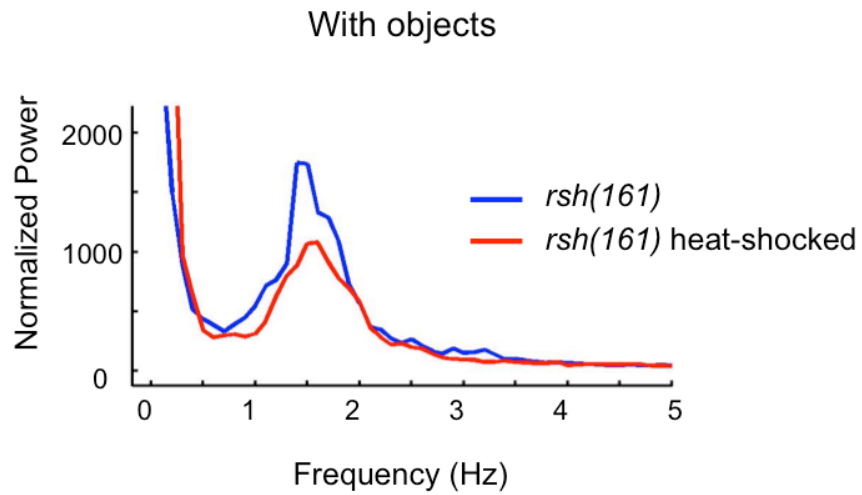
Supplemental Figure 1

**Supplemental Figure 2.** Tethered flight paradigm **A** Torques (in volts) caused by changes in wing-beat behavior for 50 s of tethered flight in a sample wild-type fly. **B** Torques behavior for 50 s of tethered flight in a sample *radish*<sup>1</sup> fly.



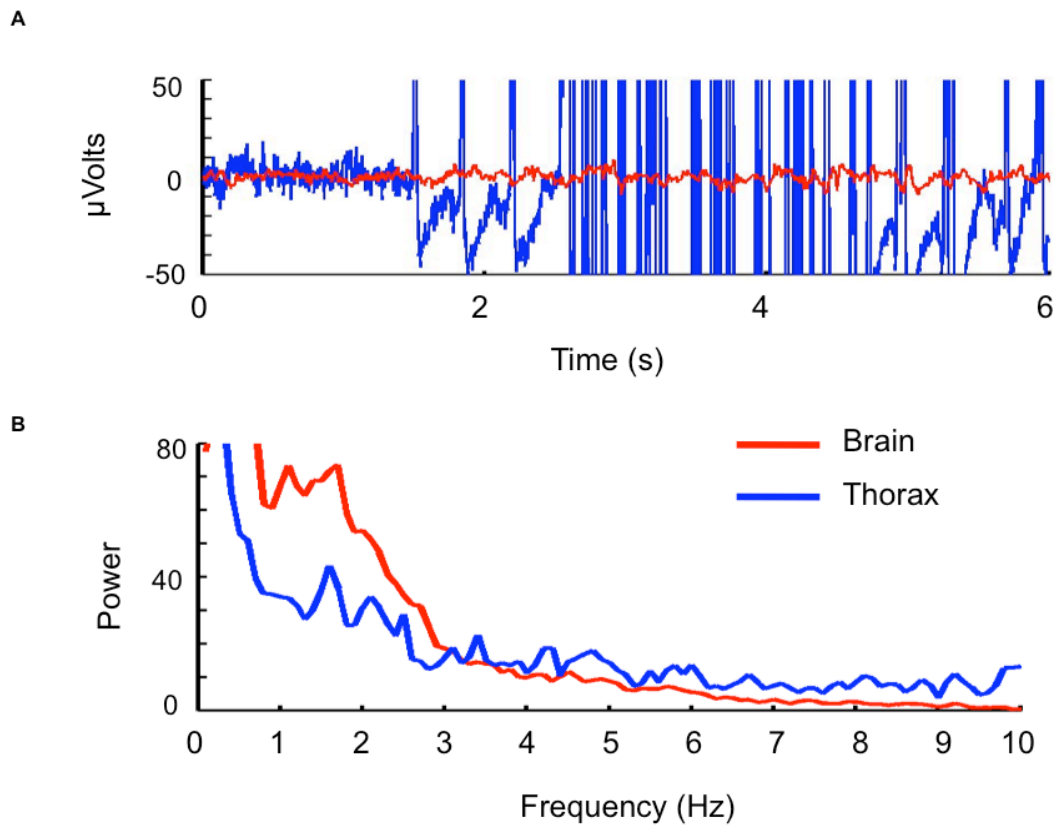
Supplemental Figure 2

**Supplemental Figure 3.** Average power spectra between 0 and 5 Hz for non heat-shocked *hs-rsh(161)* (blue line, n=4) and heat-shocked *hs-rsh(161)* (red line, n=3) 6-minute open-loop continuous flights with two distinct visual objects.



Supplemental Figure 3

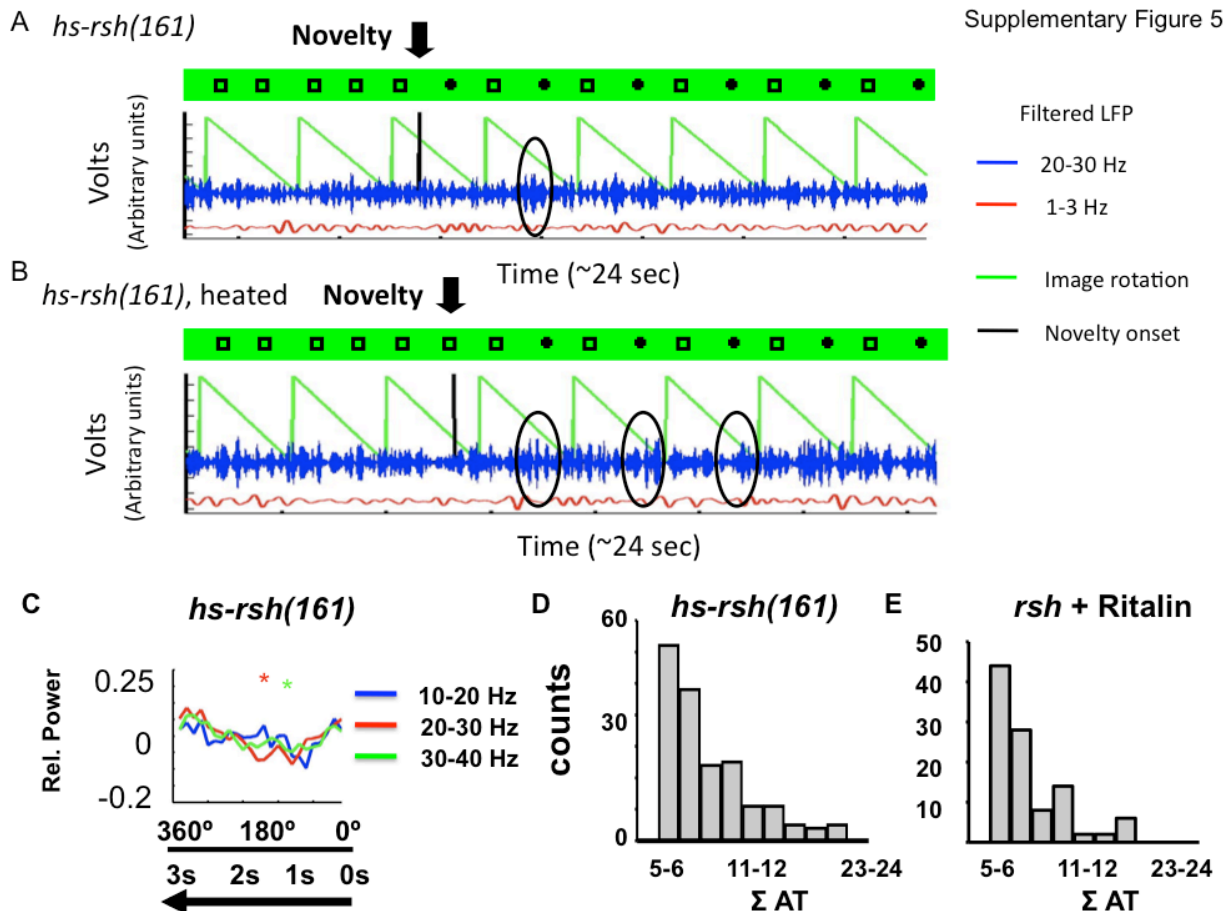
**Supplemental Figure 4.** Brain activity versus thorax activity. **A** 6s raw trace of thoracic potentials (blue) and brain LFP activity (red) in a *radish* mutant. The fly transitions to flight in the middle of the trace. **B** Z-scored spectral analysis of brain and thorax activity in a *radish* mutant, for 1-10 Hz frequencies.



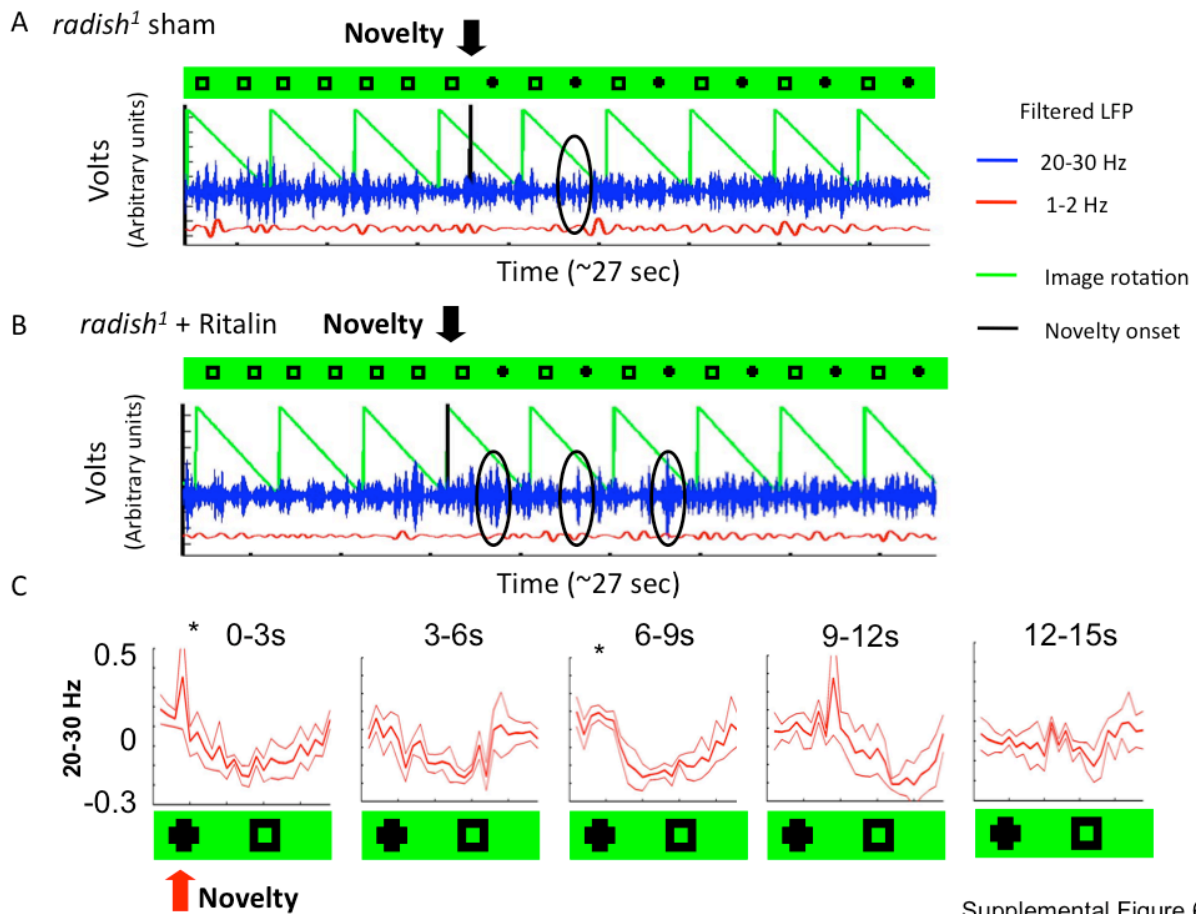
Supplemental Figure 4



**Supplemental Figure 5.** Brain responses to novelty in *hs-rsh(161)*. **A** Filtered LFP traces for a non heat-shocked *hs-rsh(161)* fly. **B** Filtered LFP traces for a (different) heat-shocked *hs-rsh(161)* fly. **C** Average novelty response for three frequency domains for 4 heat-shocked *hs-rsh(161)* flies. \* = significantly different responses ( $P < 0.05$ , by  $t$ -test) between competing objects, color coded by frequency domain. **D** Distribution of summed alternation tempos ( $\Sigma$  AT, see Methods) for 4 heat-shocked *hs-rsh(161)* flies. **E** Distribution of summed alternation tempos ( $\Sigma$  AT, see Methods) for 5 *radish* mutants treated to 0.5 mg/ml Ritalin.



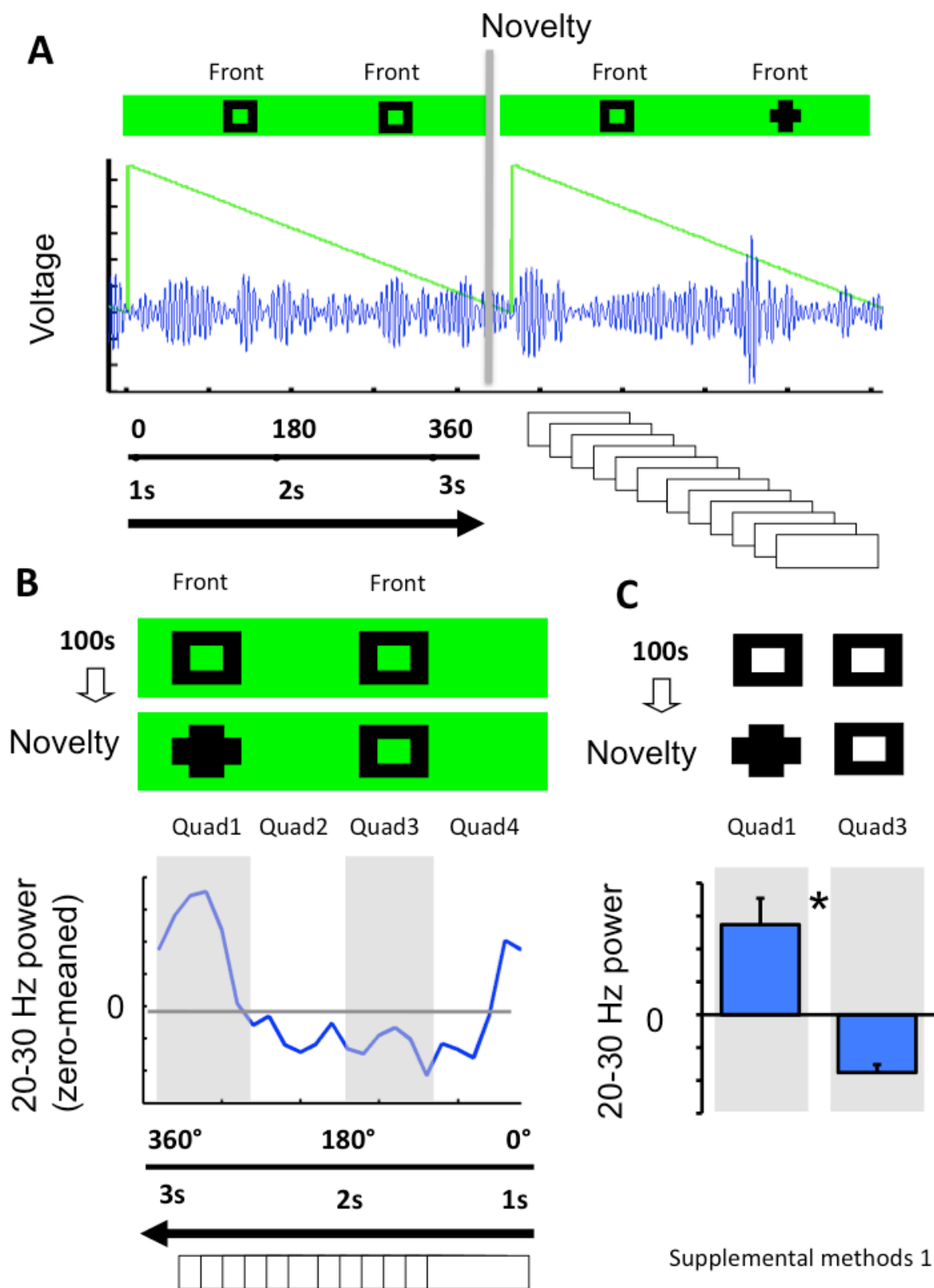
**Supplemental Figure 6.** Ritalin treatment of *radish* mutants. **A** Filtered LFP traces for a *radish* mutant. **B** Filtered LFP traces for the same fly after Ritalin feeding. **C**. Average 20-30 Hz novelty response partitioned into successive 3 s epochs for *radish* mutants fed on Ritalin (\* = significantly different responses ( $P < 0.05$ , by *t*-test) between competing objects).



### **Supplemental Methods 1:** 20-30 Hz response calculations.

**A** Three signals are recorded during an experiment: the position of the rotating visual panorama (green line), brain local field potentials (a 20-30 Hz bandpass-filtered trace is shown, blue line), and a trigger indicating a change in scene (gray bar). Each object (a square or a cross), sweeps in front of the fly at a specific time of the panorama rotation sequence, once every 3 seconds. The objects are 180° apart. 20-30 Hz power is calculated (by Fast Fourier Transform, FFT, in Matlab) for 24 overlapping windows covering the entire panorama rotation sequence (12 overlapping rectangles are shown).

**B** Average 20-30 Hz activity from the 24 FFT calculation are plotted onto the 3s rotation sequence, with image flow moving from left to right (inverted from A). For novelty experiments (100s training to squares followed by one of the squares changing to a cross), five such graphs from five separate novelty transitions are averaged to display a novelty response per fly. These data are then zero-meant and normalized in order to produce average responses ( $\pm$  s.e.m.) among flies within a strain (as in Figure 6). **C** To determine whether a strain is producing a significant response, 20-30 Hz activity is contrasted for either object (coinciding with when that object sweeps in the quadrant in front of the fly). Thus, the average 20-30 Hz activity (from B) for the quadrant representing the novel object (Quad1) is contrasted (by *t*-test) with the average 20-30 Hz activity representing the competing object (Quad3). A significant difference ( $P < 0.05$ ) indicates the 20-30 Hz selection / suppression effects characteristic of a response to visual salience effects such as novelty.





## **Supplemental Methods 2: Attention Span Calculations**

**A** Summed 20-30 Hz power is calculated for the time (1.5s) each object is in front of the fly. Thus, there are two numbers calculated for every full rotation of the panorama, (consisting of two distinct objects, a square and a cross, sweeping across in succession). A log ratio of these numbers is calculated for every rotation cycle, yielding a ratio for every successive cycle (with a total of around 130 cycles for experiment per fly. **B** The ratios are plotted in succession for the experiment (black graph). When 20-30 Hz activity was greater in response to the cross, the graph is in the blue zone; when activity was greater in response to the square, the graph is in the red zone. From this data, an “Alternation Tempo” (AT) series is calculated (red or blue numbers, corresponding to the square and the cross, respectively). These are the durations (in cycles) that the ratio is biased to one or the other object (above or below a ratio of 1) before alternating. AT durations are calculated for the entire experiment (6 are shown in b). These are then plotted as histograms in **C** The AT sequence “127531”, is indicated as a clump of alternating red and blue histograms. A clump is defined as a group of contiguous AT values where each is larger than 1, but flanked on both sides by 1’s (as in the sample clump). Clump size ( $\Sigma$ ) was calculated by summing the AT values within a clump. To quantify this behavior of the 20-30 Hz brain activity in response to competing images, we tallied clump sizes across experiments for multiple flies within a treatment or genotype. A histogram of clump sizes was then plotted, as shown in **D** Only sizes 5 or above were considered. This simple set of calculations is aimed at determining whether there are contiguous epochs of time (visualized as Alternation Tempo (AT) clumps) when the fly’s 20-30 Hz brain activity is strongly biased to one or the other object. To

test whether this potential bias is significant, we shuffled (by permutations) the original series of 20-30 Hz ratios (the black plot in b.) and then recalculated AT durations for the shuffled data. Then, clump sizes were recalculated according to the same criteria (flanked by 1's,  $\Sigma = 5$  or larger) and size distributions graphed as histograms. To test whether real data (or mutant data) were different from shuffled data, we performed a Kolmogorov-Smirnov test for distributions, with significance set at  $P < 0.05$ . Distributions that were significantly different from the shuffled set show that 20-30 Hz activity is successively biased (in time) for one or the other object. We suggest that this is a form of working memory relevant to selective attention.

